

(19) World Intellectual Property  
Organization  
International Bureau



(43) International Publication Date  
23 September 2004 (23.09.2004)

PCT

(10) International Publication Number  
**WO 2004/081038 A1**

(51) International Patent Classification<sup>7</sup>: **C07K 14/47**

(21) International Application Number:  
PCT/GB2004/001014

(22) International Filing Date: 10 March 2004 (10.03.2004)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:  
0305552.2 11 March 2003 (11.03.2003) GB  
0405190.0 8 March 2004 (08.03.2004) GB

(71) Applicant (for all designated States except US): **REGEN THERAPEUTICS PLC** [GB/GB]; Suite 406, Langham House, 29-30 Margaret Street, London W1W 8SA (GB).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **GEORGIADES, Jerzy, Alexander** [US/US]; 9615 Bayou Brook, Houston, TX 77063 (US). **POLANOWSKI, Antoni** [PL/PL]; Ul. Bystrzycka 27, PL-54-215 Wrocław (PL). **WILUSZ, Tadeusz** [PL/PL]; Ul. Tamka 2, PL-50-137 Wrocław (PL). **KRUZEL, Marian, L.** [US/US]; 13627 La Concha Lane, Houston, TX 77083 (US).

(74) Agents: **CURTIS, Philip, Anthony** et al.; A.A. Thornton & Co., 235 High Holborn, London WC1V 7LE (GB).

(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PL, PT, RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

**Published:**

- with international search report
- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: PURIFICATION OF PEPTIDES FROM COLOSTRUM

(57) Abstract: The invention relates to the purification of peptides from colostrum. The method involves the addition of an alcohol such as methanol or ethanol to the mixture in order to form an alcohol phase rich in the peptides, and a precipitate. The peptide-rich alcohol phase is subsequently recovered and subjected to further fractionation. The invention is particularly useful in the purification of colostrinin from colostrum.



**WO 2004/081038 A1**

## **PURIFICATION OF PEPTIDES FROM COLOSTRUM**

This application relates to the purification of peptides from colostrum. The invention is particularly concerned with the purification of colostrinin from  
5 colostrum.

Colostrum, or foremilk, is a viscous mammary gland secretion characterised by the presence of many elements needed by newborn mammals to develop properly. It is the first lacteal secretion post parturition and contains a high concentration of immunoglobulins (IgG, IgM and IgA) and non-specific  
10 proteins. It is replaced by mature breast milk about four to five days after birth. Compared with mature breast milk, colostrum has low sugar content, but is richer in lipids, proteins, mineral salts and immunoglobulins. It also contains various floating cells such as granular and stromal cells, neutrophils, monocyte/macrophages and lymphocytes. It is also rich in growth factors,  
15 hormones and cytokines.

Amongst the proteins present in colostrum, caseins are the most prevalent and known to form aggregates (micelles), which are similar in all mammals. Many proteins and peptides are bound to those aggregates, by weak hydrophobic and ionic forces. The resultant network of proteinaceous micelles  
20 has the ability to trap many small molecular weight compounds of differing nature, such as lipids, carbohydrates, and peptides, forming a unique homogeneous solution. The micelles help to distribute these micro-molecules relatively uniformly throughout the colostrum, and also prevent them from the formation of unwanted aggregates.

25 A number of peptides from milk with various biological activities have been reported. Some peptides exist naturally and some can be released via enzymatic proteolysis of the parent milk proteins. Of particular interest are those naturally existing peptides that are bound to casein micelles. Besides casein protein, calcium and phosphate, the micelle also contains citrate, minor ions, lipase and  
30 plasmin enzymes, and various peptides entrapped in their structure. Therefore,

the downstream purification processing of many milk-derived components recently become the most challenging effort for the dairy industry.

In general, the downstream processing of milk starts by coagulating caseins with addition of chymosin to give curds, which are then separated from the liquid, whey, after which they can be processed and matured to produce a variety of cheeses. Chymosin brakes down the kappa-casein molecules causing a global collapse of casein micelles. As a result many components of a soluble fraction of-milk become entrapped in the precipitate and disregarded. Particularly small molecular peptides with a high hydrophobic index, such as colostrinin, can be lost.

The original purification protocol for obtaining colostrinin (Janusz et al.), consists of pH-dependent casein precipitation followed by various chromatographic steps, including ion exchange, affinity and molecular sieving, combined with ammonium sulfate precipitation. Although, this method is reproducible, it is laborious and difficult to scale-up for the industrial applications. Since then numerous protocols, utilizing membrane filtration, have been developed for recovery of low molecular weight peptides from milk and colostrum, but all of these have had shortcomings.

Tangential flow filtration is used in one standard protocol for separating milk components in the dairy industry. For example, in the US Patent 6,268,487 entitled *Purification of Biologically Active Peptides from Milk* by Kutzko et al., a method for separation of milk components by tangential flow filtration is disclosed.

Also, an ultrafiltration is proposed by Roger et al., in a US patent 4,485,040, entitled: *Process for Obtaining an  $\alpha$ -Lactalbumin Enriched Product from Whey, and Uses Thereof* , to separate milk components on a membrane having cut-off greater than 5,000 (e.g. 50,000) at a pH between 6.3 and 7 (e.g. 6.6) and a temperature between 30°C and 60°C.

The use of ultrafiltration is also described in US Patent 4,816,563 Wilson et al., entitled: *Process for Obtaining Transfer Factor from Colostrum, Transfer Factor so Obtained and Use Thereof*. In fact, filtration of milk or colostrum is a

standard procedure in the dairy industry today. Wilson et al., describes the use of an agent to prepare a transfer factor from colostrum. Suitable agents are said to include alcohols, ketones and polyethylene glycol. There is, however, no description of the recovery of colostrinin from colostrum.

5       US Patent 5,216,129 discloses a process for obtaining kappa-caseinoglycomacropeptide from a whey product concentrated in proteins. The process involves the use of ethanol at a concentration of 5% to 25% based on the volume of the solution. The whey product is not colostrinin.

10       UK Patent 1,438,008 discloses a process for the extraction of a specific octapeptide from frog skin using alcohol.

JP520062796 discloses a method of extracting cyclic peptides from the root of *Ribia akane* Nakai using, *inter alia*, an alcohol.

15       In J Pharm Pharmaceut Sci, Vol 5, 2002. MEC Lutsiak et al, "Analysis of peptide and lipopeptide content in liposomes", p 279-284, there is disclosed a method for extracting peptides from liposomes.

In Journal of Antibiotics, Vol XL, 1987, E Meyers et al, "Xylocandin: a new complex of antifungal peptides I. Taxonomy, isolation and biological activity", p 1515-1519, there is disclosed method of extracting peptides from the bacteria *pseudomonas cepacia*.

20       In Acta Endocrinologica, vol 111, 1986, WF Blum et al, "Isolation and partial characterisation of six somatomedin-like peptides from human plasma Cohn fraction IV", p 271-284, there is disclosed the extraction of somatomedin-like peptides from human plasma using ethanol.

25       In J Dairy Research, vol 54, 1987, DS Horne, "Ethanol stability of Casein micelles - a hypothesis concerning the role of calcium phosphate", p 389-395, there is disclosed a hypothetical elaboration on casein micelles structure and calcium release.

30       In Ir. J. Fd. Sci. Technol., vol 9, 1985, MM Hewedi et al, "Recovery of milk protein by ethanol precipitation", p 11-13, there is disclosed a method of precipitating milk proteins using ethanol.

Colostrinin, also known as colostrinine, proline-rich polypeptide or PRP, was first isolated in 1974 (Janusz et al, FEBS Lett., 49, 276-279) from ovine colostrum.

Certain therapeutic uses of colostrinin, particularly in the treatment of  
5 Alzheimer's disease, were described in WO98/14473, the contents of which are incorporated herein by reference. In this patent application, the physical characteristics of colostrinin, as determinable at the time, were described. Although the physical characteristics were correct, the understanding of colostrinin has moved on since this application was filed. WO98/14473 also  
10 described a method for extracting colostrinin from raw colostrum, which is often referred to as the "Janusz" method. This method is presently the principle method of extracting colostrinin from colostrum. It has the disadvantages that the industrial scale up is difficult to obtain and yields from the method are low.

WO00/75173, the contents of which are incorporated herein by reference,  
15 describes a number of peptides found in colostrinin. WO02/46211, the contents of which are incorporated herein by reference, describes a number of other peptides which can be found in colostrinin.

Of considerable interest in colostrinin is the presence of various polypeptides that can be isolated only from colostrum, not mature milk. During  
20 the days following parturition, the concentration of colostrinin in a mammary gland secretion precipitously diminishes through the end of the third day after delivery. Such a short lifetime for some of the colostrinin peptides indicates their important role in early development of infant's immune system and the protection of newborns against environmental shocks.

25 Colostrinin has more recently been discovered to exist in two forms; free and aggregated (bound). It is thought that free forms of colostrinin are required to protect the newborn mammal from oxidative stress, which appears immediately after birth. The bound or aggregated forms are designed to maintain this function for prolonged periods of time after birth. The bound form of  
30 colostrinin, in addition, is thought to participate in the development and/or protection of different organs and systems. This takes place when oxidative

stress problems subside, and free colostrinin concentrations begin to decrease. The bound form of colostrinin is slowly released into the body fluids to modulate physiological functions when the free form is exhausted. This model finds support in studies on the gradual disappearance of specific colostrinin peptides from colostrum after parturition.

The colostrinin complex is now believed to consist of at least five subgroups of peptides; each subgroup has its own characteristic hydrophobic pattern. Evidence suggests that these peptides have a tendency to form aggregates, due to the presence of specially arranged non-polar, polar, aromatic, positively- and negatively-charged amino acids. Furthermore, the amino acid compositions of the peptides and their hydrophobic character further suggest this aggregating ability.

From the presently available information, we have found that colostrinin peptides have the best biological activity when they are present in their native form. When they are purified, they start to interact with each other forming non-covalently bound complexes with apparently weaker biological activities. It is believed that colostrinin is a mixture of more than 62 separate peptides, derived from precursor proteins, such as annexin, beta-casein, a hypothetical beta-casein homologue and others with no homology to any specific protein in the current GenBank database.

It is an object of the invention to develop a new method for extracting and recovering peptides from fluids containing higher molecular weight materials. It is a specific object of the invention to develop a method of purifying colostrinin from raw colostrum, so that the colostrinin can be produced in high yield in a form in which it will be biologically active and substantially pure. The purification method should yield the majority of the colostrinin peptides, both free and bound or aggregated. Purified colostrinin, free of contaminants and self-aggregates, will have more active principles per unit weight. A further object of the invention is to develop a purification protocol that will provide a consistent set of peptides characterised by specific biological activities.

This invention is based on the unexpected discovery that a simple extraction method can be used to extract colostrinin at a high degree of purity from colostrum.

According to one aspect of the invention there is provided a method for recovering peptides from mammalian colostrum said method comprising: mixing the colostrum with an alcohol to form an alcohol phase and a precipitate, the alcohol phase including the alcohol and at least some of the peptides to be recovered; separating the alcohol phase from the precipitate; and recovering the alcohol phase.

The invention is particularly applicable to the purification of colostrinin from colostrum, and the following description is largely directed to this specific application.

The alcohol may be any alcohol capable of forming an alcohol phase containing colostrinin peptides when mixed with colostrum, and capable of precipitating unwanted higher molecular weight materials.

The alcohol may be linear or branched, and may contain one or more hydroxyl groups; one hydroxyl group is preferred. Preferably, the alcohol contains 1 to 5 carbon atoms, more preferably 1 to 4 carbon atoms, most preferably 1 to 3 carbon atoms. We have found that the best results are obtained with methanol or ethanol.

The alcohol is desirably added to the colostrum in a concentrated form: preferably the alcohol is at least 80% pure, more preferably at least 95% pure, and most preferably substantially 100% pure.

The amount of alcohol added to the colostrum is preferably such as to provide an alcohol concentration in the total composition of 40% (v/v) to 80% (v/v), more preferably 50% (v/v) to 70% (v/v), still more preferably 55% (v/v) to 65% (v/v) and most preferably substantially 60% (v/v).

The step of mixing the alcohol with the raw colostrum (which is known as "extraction"), is preferably carried out with stirring at room temperature for 10 to 30 minutes. This forms a precipitate containing the caseins and other proteins, while the colostrinin remains in solution in the alcohol phase. It is an important,

and unexpected, feature of the invention that the colostrinin peptides substantially all remain in solution in the alcohol, while the other components of the colostrum are precipitated.

The precipitate may then be separated from the alcohol phase by any  
5 conventional means, preferably by centrifugation at about 15,000 g, and the alcohol phase containing the colostrum is recovered. The colostrinin is then separated from the alcohol, preferably by evaporation, to form a colostrinin-rich phase (which is largely an aqueous solution of colostrinin) which is recovered.

The step of separating the alcohol from the colostrinin peptides can be  
10 carried out by evaporation or extensive ultrafiltration, combined with water or buffer exchange, but the evaporation is preferred. The evaporation can be carried out at a temperature typically in the range of 10°C to 50°C. A temperature of about 30°C is preferred. The evaporation or filtration may be carried out over a time period of, for example, 30 minutes to 12 hours or more (e.g. overnight).  
15 Although the ultrafiltration is a viable option, the recovery is higher with evaporation. It may be desirable to eliminate alcohol by evaporation in a vacuum. After removal of the alcohol, sufficient water may be added to the colostrinin-rich phase to bring-up the volume of working solution to about initial volume.

In a preferred embodiment, the precipitate from the alcohol-extraction step  
20 is subsequently washed with a further quantity of alcohol, and the alcohol phase is again separated and recovered. This second recovered alcohol phase may contain some colostrinin peptides which were carried into the precipitate in the first extraction. The second recovered alcohol phase may be added directly to the alcohol phase recovered from the first extraction, or some or all of the alcohol  
25 may be removed first. After the alcohol has been removed, the remnants could be added directly to the colostrinin-rich phase.

Optionally, following removal of the alcohol, a further centrifugation step may be performed in order to separate any precipitate formed during the alcohol removal from the remaining solubilized fraction.

30 In a particularly advantageous embodiment of the invention, a precipitation agent is added either to the alcohol phase, or, preferably, to the colostrinin-rich



phase (preferably after any additional centrifugation step). The purpose of the precipitation agent is to induce precipitation of the colostrinin peptides. We have found that ammonium sulfate is particularly suitable as the precipitation agent, but other materials which can fulfil the same purpose could be used instead, or in addition. The most effective precipitation agents are salts with multiple charged anions such as sulphate, phosphate, and citrate. As far as cations are concerned, monovalent ions should be used with  $\text{NH}_4^+$  being preferred to  $\text{K}^+$ , and  $\text{K}^+$  being preferred to  $\text{Na}^+$ . Typical precipitation agents are sodium, potassium and ammonium sulphates, phosphates and citrates, but ammonium sulphate is most preferred. The precipitation agent may instead be an organic polymer, such as polyethylene glycol. Precipitation with a precipitation agent further lowers the casein contamination and further purifies the colostrinin peptide fraction, enabling the formation of a consistent pool of essentially IgG-free peptides to be isolated in approximately two days. This process makes possible the rapid isolation of colostrinin peptides.

The precipitation agent is desirably added in an amount suitable to provide 30% (w/v) to 80% (w/v) saturation, preferably 40% (w/v) to 60% (w/v) saturation, and most preferably substantially 50% (w/v) saturation. The saturation percentages refer to saturation of the precipitation agent, such as ammonium sulfate, in liquid. The precipitation agent is preferably added as a saturated solution (i.e. 100% (w/v) saturation) in water.

However, the precipitation agent may also be added in crystalline form, in small amounts, followed by a vigorous stirring to avoid local saturation, for about 30 minutes for example, after each addition. This may be continued until the entire precipitation agent has been dissolved. This technique may take several hours, therefore is less preferred.

The addition of the precipitation agent causes the colostrinin peptides to be precipitated from the solution. When the addition is complete, the mixture is preferably left for a period of time, typically 30 minutes, possibly with stirring, or, preferably, with rocking.

At this stage, the mixture comprises an aqueous liquid phase and a precipitate. The liquid phase contains the precipitation agent dissolved therein, and, possibly, some of the higher molecular weight material which was not precipitated during the alcohol extraction step. The precipitate contains colostrinin peptides. The liquid phase is subsequently separated from the precipitate, preferably by centrifugation at about 15,000 g centrifugal force, and the precipitate is recovered.

The recovered precipitate is preferably dissolved in water, using the minimum amount of water necessary, followed by dialyzing, typically in 0.01M PBS (phosphate buffered saline) or water. The dialysis step is carried out to remove excess ions, although some ions of the precipitation agent ions may remain. The dialysed solution may then be clarified by centrifugation, then lyophilised. The final material contains separated colostrinin peptides, which may be stored at -20°C for future use in the preparation of a pharmaceutical, nutraceutical and other compositions. These may be made by conventional means.

Various quality control tests may be carried out, to ensure that the final material is suitable for use in the preparation of a medicament. Such tests are known to those skilled in the art, but may involve SDS PAGE; isoelectrofocusing profile; amino acid analysis; determining the antigenic composition of colostrinin by means of the ELISA method using monospecific antibodies; determining the capability to induce cytokines such as interferon gamma (IFN- $\gamma$ ); tumour necrosis factor (TNF) and determining the antioxidant properties. The final material should not contain any high molecular weight proteins. If the quality control tests indicate the presence of impurities, the final material may need to be further purified. There are several methods by which the impurities can be removed from the final material, such as size exclusion chromatography, hydroxyapatite chromatography, reverse phase chromatography, ultrafiltration and fractionation by perchloric acid.

An example of size exclusion chromatography involves the following steps. The material, from which the impurities have to be removed, is

reconstituted in 100mM phosphate buffer, pH 7.2 (concentration - 2.0 mg/ml). Ten millilitres of freshly reconstituted material is applied on a column (2.5 cm diameter x 90 cm length) packed with Bio-Gel P30 (BioRad) and equilibrated with the same buffer. The chromatography is developed overnight at a flow rate of 20 ml/hour. The samples (4 ml) are collected and the final pool of material corresponding to low molecular weight proteins and peptides (>18K) is made according to SDS PAGE distribution.

An example of hydroxyapatite chromatography involves the following steps. The material, from which the impurities have to be removed, is reconstituted in 10mM phosphate buffer, pH 6.5 (concentration - 2.0 mg/ml). Ten millilitres of freshly reconstituted material is applied on a column (0.7 cm diameter x 6 cm length) packed with Bio-Gel HTP (Bio-Rad) and equilibrated with the same buffer. The chromatography is developed at a flow rate of 0.25 ml/min. The column is washed with equilibrating buffer to remove all unbound material. Subsequently, the elution is performed with a linear gradient of phosphate buffer (10 mM to 500 mM). The samples (4 ml) are collected and the final pool of material corresponding to low molecular weight proteins and peptides (>18K) is made according to SDS PAGE distribution.

An example of reversed-phase High Performance Liquid Chromatography involves the following steps. The material, from which the impurities have to be removed, is reconstituted in 0.1% (v/v) of trifluoroacetic acid (TFA) at a concentration - 2.0 mg/ml. Five millilitres of freshly reconstituted material is applied on a column (8 mm diameter x 150 mm length) packed with 10 $\mu$ m Nucleosil 100 C18 (Knauer) and equilibrated with 20% acetonitrile in 0.1% TAF. The chromatography is developed at a flow rate of 1.0 ml/min. The column is washed with equilibrating buffer to remove all unbound material. Subsequently, the elution is performed with 100% acetonitrile in 0.1% TFA. The samples (2 ml) are collected and the final pool of material corresponding to low molecular weight proteins and peptides (>18K) is made according to SDS PAGE distribution.

An example of perchloric acid fractionation involves the following steps. The material, from which the impurities have to be removed, is reconstituted in

10mM phosphate buffer, pH 7.2 (concentration - 2.0 mg/ml). Perchloric acid (0.45 M HClO<sub>4</sub>) is added up to the concentration of 0.15 M and the mixture is stirred at room temperature for 1 hour. The resulting precipitate is removed by centrifugation and the supernatant, containing purified CLN is adjusted to pH 7.0  
5 with 1M KOH and left overnight. The crystals of KClO<sub>4</sub> are removed by filter filtration and supernatant is dialyzed against water overnight. In another embodiment of the invention perchloric acid fractionation is use as a substitute of the ammonium sulphate precipitation step.

An example of ultrafiltration involves the following steps. The material,  
10 from which the impurities have to be removed, is reconstituted in 10mM phosphate buffer, pH 7.2 (concentration - 2.0 mg/ml). Ultrafiltration is carried out in Centricon Plus Ultracell PL-10 or Hollow fiber Amicon H1P10-20 using standard ultrafiltration protocols.

As mentioned above, we have found that colostrum contains a  
15 proteinaceous fraction in the form of micelles. It has been observed that colostrinin can be recovered at higher yield when the raw colostrum is pre-treated with a micelle-breaking material, followed by a micelle-restoring agent, at various pH levels, prior to extraction with alcohol. Therefore, in an advantageous embodiment of the invention, the raw colostrum is pre-conditioned with a micelle-  
20 breaking material, which induces collapse of the micelles, then with a micelle-restoring agent, which is believed to disintegrate casein structures in colostrum, releasing more colostrinin peptides.

Preferably the micelle-breaking material is a calcium-chelating agent, such as EDTA (ethylene-diamine-N,N,N',N'-tetraacetic acid), which also binds Mg<sup>+</sup>, or  
25 EGTA (ethylene glycol-O,-O'-bis(2-amino-ethyl)-N,N,N',N'-tetraacetic acid) with similar characteristics. The purpose of this is to release at least some of, preferably a majority of, the peptides entrapped between casein micelles.

The micelle-breaking material is preferably added in an amount to produce a solution that contains a concentration of micelle-breaking material in solution of  
30 25-250 mM. The mixture is stirred and left to stand for a period of 10 - 30 minutes. The alcohol may be added immediately after addition of the micelle-

dissolving material, or, in a more preferred embodiment, the addition of the micelle-breaking material is followed by addition of a micelle-restoring material, prior to the addition of the alcohol. The micelle-restoring material is preferably a source of  $\text{Ca}^{2+}$  ions such as  $\text{CaCl}_2$ . The micelle-restoring material has the capacity to re-form micelles by providing calcium ions essential for micelle formation (recalcification).

In a preferred embodiment, the colostrum is conditioned to a pH from 5.5 to 8.0, more preferably 5.5-7.5 prior to the addition of the micelle-breaking material, the micelle-restoring material or the alcohol. Most preferably, the colostrum is conditioned to a pH from substantially 6.5-7.5. Still more preferably, the colostrum is conditioned to a pH from substantially 7.0 to 7.5, most preferably substantially 7.4. The pH conditioning may be achieved by addition of acid or alkali under the control of a pH meter.

A consistent pool of essentially IgG-free peptides can be isolated in a high yield using the method according to the invention. By simultaneously neutralising hydrophobic and ionic forces, we can obtain an optimum physiological combination of peptides. The yield of colostrinin peptides produced by extraction with methanol, according to the present invention, is generally around 500-600 mg per litre colostrum. Using pretreatment with EDTA and calcium chloride, the yield increased up to around 4000-5000 mg per litre colostrum. This compares with a prior art yield of around 200-300 mg per litre colostrum using the Janusz method described in WO98/14473.

The invention may be used in the separation from colostrum of any of the peptides known to be present in colostrum, specifically:

(i) the peptides disclosed in WO00/75173, i.e., LQTPQPLLQVMMEPQGD (SEQ ID 1); MPQNFYKLPQM (SEQ ID 2); VLEMKFPPPPQETVT (SEQ ID 3); LKPFPKLKVEVFPFP (SEQ ID 4); SEQP (SEQ ID 5); DKE (SEQ ID 6); DPPPPQS (SEQ ID 7); LNF (SEQ ID 8); VLPPNVG (SEQ ID 9); KYKLQPE (SEQ ID 10); SEEMP (SEQ ID 11); DSQPPV (SEQ ID 12); FPPPK (SEQ ID 13); VVMEV (SEQ ID 14); DLEMPVLPVEFPFPV (SEQ ID 15); LFFFLPVVNVLP (SEQ ID 16); MQPPPLP (SEQ ID 17); DQPPDVEKPDLPFQVQS (SEQ ID 18);

VYPFTGPIPN (SEQ ID 19); SLPQNILPL (SEQ ID 20); TQTPVVVPPF (SEQ ID 21); LQPEIMGVPKVKETMVPK (SEQ ID 22); HKEMPFPKYPVEPFTESQ (SEQ ID 23); SLTLTDVEKLHLPLPLVQ (SEQ ID 24); SWMHQPP (SEQ ID 25); QPLPPTVMFP (SEQ ID 26); MHQPPQPLPPTVMFP (SEQ ID 27); PQSVLS (SEQ ID 28); LSQPKVLPVPQKAVPQRDMPIQ (SEQ ID 29); AFLLYQE (SEQ ID 30); FLLYQEPVLGPVR (SEQ ID 31); RGPFPILV (SEQ ID 32); or ATFNRYQDDHGEEILKSL (SEQ ID 33).

(ii) the peptides disclosed in WO02/46211, i.e., LVYPFTGPIPN SLPQNILP (SEQ. ID 34); MIVVRL LQNEVPE (SEQ. ID 35); SLSQSKVLPV (SEQ. ID 36); LQTQTPVV (SEQ. ID 37); EMPFPKY (SEQ. ID 38); PVEPFT (SEQ. ID 39); VPPFLQ (SEQ. ID 40); PMFLQ (SEQ. ID 41); EHM FV (SEQ. ID 42); TDRD (SEQ. ID 43); VQPT (SEQ. ID 44); PKVK (SEQ. ID 45); DDDE (SEQ. ID 46); TEEV (SEQ. ID 47); YQQE (SEQ. ID 48); FPPQ (SEQ. ID 49); GFGI (SEQ. ID 50); LQS (SEQ. ID 51); VVV (SEQ. ID 52); GGK (SEQ. ID 53); DMV (SEQ. ID 54); ESQ (SEQ. ID 55); GRV (SEQ. ID 56); VEE (SEQ. ID 57); IGN (SEQ. ID 58); FFQ (SEQ. ID 59); RMF (SEQ. ID 60); FPP (SEQ. ID 61); MHH (SEQ. ID 62); NTE (SEQ. ID 63).

(iii) the nonapeptides disclosed in WO98/14473, i.e., VESYVPLFP (SEQ. ID 64).

Thus the invention may be used in processes to isolate any of the peptides identified by SEQ. ID 1-64, either individually, in a selected group, or all together.

The invention may be applied to any mammalian colostrum, although ovine, bovine or human colostrum are most commonly used. Furthermore, the invention has more general application beyond the separation of colostrinin peptides from colostrum. It is believed that the present invention has general applicability to the separation of small peptides from a mixture of the peptides with protein, lipids and other biological materials.

Furthermore, the starting point for the separation process according to the invention need not be colostrum as obtained directly from the mammal. It will be appreciated that the colostrum may be treated prior to being subjected to the

separation method according to the invention, for example it may be defatted. Thus, the invention may be applied, with advantage to a derivative of colostrum, provided that the derivative still contains the colostrinin.

The method according to the invention may be applied, with advantage, to  
5 any body fluid, including cerebrospinal fluid, saliva, blood, ascitis or urine. The invention is particularly applicable to separations involving peptides in dairy fluids.

It will, therefore, be appreciated that the techniques described above may have wider application to the general separation of peptides from fluids  
10 containing higher molecular weight components including, but not limited to, proteins, lipids, carbohydrates and/or nucleic acids. In general, the fluid will be in the form of an aqueous liquid, having each of the peptides and other components either suspended in or dissolved in the liquid. Thus, according to another aspect of the present invention there is provided a method for recovering peptides from  
15 a fluid containing said peptides in combination with higher molecular weight materials, such as proteins, lipids, carbohydrates and/or nucleic acids, said method comprising: mixing the fluid with an alcohol to form an alcohol phase and a precipitate, the alcohol phase including the alcohol and at least some of the peptides, and the precipitate including at least some of the higher molecular  
20 weight materials; separating the alcohol phase from the precipitate; and recovering the alcohol phase.

References to % compositions herein relate to the percentage of the material in parts by weight, unless stated otherwise.

Reference is now made to the accompanying drawings, in which:

25 Figure 1 is a chart summarising the extraction/purification protocol for colostrinin; and

Figure 2 shows the results of the SDS page analysis described below.

The invention will be further described with reference to the following examples.

Purification Protocols - For each comparative example, 10 ml of pooled colostrum was utilised. The colostrum was collected from sheep at different times postpartum and frozen until experiment. The protocol utilised in the following examples was identical, with the exception of initial conditioning of raw material either by pH or EDTA, as indicated below.

Example 1 – 10ml of pooled raw colostrum (pH ~ 6.5).

Example 2 – 10ml of pooled raw colostrum (pH ~ 6.5) conditioned with 25mM EDTA, followed by addition of 50 mM  $\text{CaCl}_2$  (recalcification).

Example 3 – 10ml pooled colostrum conditioned at pH 7.4.

Example 4 – 10ml pooled colostrum conditioned at pH 7.4 with 25mM EDTA, followed by addition of 50 mM  $\text{CaCl}_2$  (recalcification).

The same source of pooled raw colostrum (10ml aliquots) was used to condition initial material at specific pH or EDTA/ $\text{CaCl}_2$  treatment for 1 hour prior to addition of methanol. 100% methanol was added, up to a final concentration of 60% to each colostrum sample. The mixture was stirred for 30 minutes at room temperature, and was then centrifuged at 15,000 g. The supernatant was removed, and the alcohol allowed to evaporate from the supernatant in a dryer, until the volume of the supernatant was reduced to around 8.0 ml (this took around 30-60 minutes). An equal amount of saturated ammonium sulphate (100%) was added to the supernatant to obtain a final concentration of 50% ammonium sulfate. The supernatant/ammonium sulfate mixture was then rocked gently overnight at 4°C, allowing colostrinin peptides to precipitate. The preparation was centrifuged at 15,000 g to form a pellet, containing the colostrinin peptides. The pellet was dissolved in distilled water and dialyzed against 0.01M PBS (phosphate buffered saline) overnight. The colostrinin peptides prepared by this method can be analysed in various tests and stored at -20°C.



*Protein Recovery Assay* - The protein concentration can be taken by reading the optical density of the solution at 280nm prior to freezing. The protein recovery results are shown in Table 1, below.

5 Table 1

Example/Sample	OD 280nm	Volume (ml)	Yield (mg)
1- Raw Colostrum pH 6.5	0.83	2.0	1.66
2- Colostrum/EDTA/ pH 6.5	0.86	2.0	1.72
3- Colostrum pH 7.4	3.97	2.0	7.94
4- Colostrum/EDTA/pH 7.4	9.44	2.0	18.88

The results indicate that protein recovery from the original 10ml aliquots of colostrum is highly dependent on pH. These yields are higher than using the Janusz method and they are obtained using an easier and faster process.

*Antigen Recognition Assay* – The antigenic profile of the final material was determined by the ELISA method based on the antibodies prepared against nine synthetic peptides, for which sequences have been identified previously and are listed in Table 2.

15

Table 2

Antigen Class	Sequence	Titer
A-1	LQTPQPLLQVMMEPOGD	>25,600
A-3	VLEMKFPPPPQETVT	>25,600
B-8	DLEMPVLPVEFPFV	>25,600
B-9	LFFFLPVVNVL	>25,600
C-2	SLPQNILPL	>25,600
C-11	LSQPKVLPVPQKAVPQDMPQ	>25,600
D-1	ATFNRYQDDHGEEILKSL	>25,600
LF	KCRRWEWRMKKLGAPSITCVRRAF	>25,600
IgG 2	Whole molecule	0

The antigen classes A, B, C and D are explained in WO00/75173. LF is a fragment of lactoferrin, having antibacterial property, IgG-2 antibody fragments serve as an indicator of colostrinin purity.

Briefly, 96 well ELISA plates were coated with samples of colostrinin recovered according to protocol 1-4 (approximately 10 micrograms of protein per 1ml of 0.1 M bicarbonate buffer (pH 9.0). The plates were incubated at 37°C for 3 hours, washed with coupling buffer, and blocked with a standard solution of bovine serum albumin (BSA). 50 microlitres of diluent BSA (0.75% solution) was pipetted into each well and 50 microlitres of serum sample (Protein A purified rabbit antibody against specific antigen), diluted according to the initial titre, were added to column A of each row. 1:2 serial dilutions were made, moving down the plates. The plates were incubated, covered, for 60 minutes at room temperature, then washed 4 times with PBS solution. A volume of 50 microlitres of goat anti-rabbit IgG-horseradish peroxidase conjugated antibodies (1:1000 dilution in BSA) was pipetted into each well and incubated for 60 minutes at room temperature. The plates were then washed 4 times with PBS wash solution and 50 microlitres of substrate (2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid-diammonium salt) was added to each well, and incubated for 2 minutes at room temperature. The colorimetric reaction was stopped by adding 50 microlitres of 1% SDS to each well, and the plates were read on a Dynatech plate reader (at 405nm).

The results of these experiments (mean titres) are shown in Table 3 below. The values are in Units/ml  $\times 10^3$ . A Unit is defined as a final dilution for the recognition of antigen. It is clear that the recovery of individual peptides for which the final materials were tested (A-1; A-3; B-8; B-9; C-2; C-11; D-1; Lf and IgG-2) was highest for the extractions made at pH 7.4 (Experiment 3 and 4). In both cases the antigens were generally recognized to the highest dilutions tested.

Table 3

Experiment	A-1	A-3	B-8	B-9	C-2	C-11	D-1	Lf	IgG-2	Control	Experiment
1	6400	6400	12800	6400	25600	25600	3200	1600	0	0	1
2	12800	12800	12800	6400	25600	25600	3200	6400	0	0	2
3	25600	12800	25600	25600	25600	25600	25600	25600	0	0	3
4	25600	25600	25600	25600	25600	25600	12800	12800	0	0	4

*Amino acid analysis* – The amino acid composition of Colostrinin purified according to present invention confirms high content of proline (~20%) and acidic amino acids. Table 4 shows high degree similarity in amino acid composition between Colostrinin described previously and this present invention.

Table 4 Amino acid analysis of CLN

Amino Acid	Colostrinin		
	Original *	Original**	Novel***
Asp/Asn	3.42	3.05	2.80
Ser	5.66	5.70	5.05
Glu/Gln	15.48	15.29	15.77
Gly	3.15	3.13	3.03
His	2.54	2.45	2.14
Arg	2.32	2.45	3.34
Thr	5.73	5.69	5.30
Ala	2.78	2.99	2.13
Pro	21.07	20.92	22.50
Tyr	1.36	1.55	1.54
Val	9.27	10.34	11.10
Met	3.33	3.08	1.70
Lys	5.30	5.19	4.93
Ile	3.17	2.92	3.42
Leu	11.04	11.27	10.47
Phe	4.38	4.42	4.77

\* - according to M. Janusz *et al.* ref. (FEBS LETTERS 1974, 49, 276-279)

\*\* - according to the average results obtained in further work using the Janusz *et al.* protocol

\*\*\* - according to the method according to the invention

*Biological assays:* The potency of CLN produced according to this novel protocol has been tested in comparison with classical method and proved to be similar in the following activities: 1) reduction of 4HNE-protein adducts; 2) reduction of intracellular levels of ROS; 3) inhibition of 4HNE-mediated glutathione depletion; and 4) inhibition of 4HNE-induced activation of c-Jun NH2-terminal kinases.

SDS PAGE analysis. 1D 15% polyacrylamide gel electrophoresis was carried out under reducing conditions. Staining with 1% Coomassie Blue shows striking similarities between final preparations of CLN obtained according to present invention and classical method.

Figure 2 shows the results of the SDS PAGE analysis. Samples were rehydrated, assayed for protein contents and equal amount of protein was subjected to 1D 15% gel SDS PAGE followed by staining with Coomassie Blue. In the drawing STD represents molecular weight markers; 1 represents colostrinin according to Janusz et al.; 2 represents colostrinin according to the present invention; and 3 represents colostrinin according to the present invention modified by EDTA/CaCl<sub>2</sub>. As per SDS PAGE analyses, colostrinin obtained according to the present invention is substantially identical to the referenced material (Janusz method). Also the modification with EDTA/CaCl<sub>2</sub> provides similar material, although, there is an increase in higher molecular weight protein composition.

It will be appreciated that the invention may be modified. For example extraction agents other than alcohol may be utilised for the extraction. Some success has been achieved with ketones, such as acetone. Also, the ammonium sulphate precipitation step can be substituted with the perchloric acid fractionation as described previously. Whilst the starting point for the method is described as raw colostrum or defatted colostrum, it will be appreciated that other forms of colostrum or partially purified colostrum may be utilised as the starting point. For example, the method of the present invention could utilise and further purify colostrinin peptides that have previously been purified by the known prior art methods, such as the Janusz method.

**CLAIMS**

1. A method for recovering peptides from mammalian colostrum, or a derivative of colostrum, said method comprising: mixing the colostrum with an alcohol to form an alcohol phase and a precipitate, the alcohol phase including the alcohol and at least some of the peptides to be recovered; separating the alcohol phase from the precipitate; and recovering the alcohol phase.

2. A method according to claim 1, wherein the alcohol is methanol or ethanol

3. A method according to claim 1 or 2, further comprising the step of removing at least some of the alcohol from the recovered alcohol phase to leave a concentrated peptide phase, and recovering the concentrated peptide phase.

4. A method according to claim 1, 2 or 3, further comprising dissolving a precipitation agent in either the recovered alcohol phase or in the concentrated peptide phase, said precipitation agent being capable of inducing precipitation of at least some of the peptides.

5. A method according to claim 3 or 4, wherein the precipitation agent is ammonium sulfate.

6. A method according to any preceding claim, further comprising adding EDTA to the colostrum prior to adding the alcohol.

7. A method according to any one of claims 1 to 5, wherein the mixture contains protein micelles to which the peptides are bound, said method further comprising the step of adding a micelle-breaking material to the fluid, prior to the addition of the alcohol, the micelle-breaking material being capable of inducing break up of the micelles in the fluid.

8. A method according to claim 7, wherein the micelle-breaking material is EDTA or EGTA.

9. A method according to any preceding claim, further comprising adding  
5  $\text{CaCl}_2$  to the mixture prior to adding the alcohol.

10. A method according to claim 7 or 8, further comprising adding a micelle-restoring material to the fluid, prior to adding the alcohol, in order to re-form the protein micelles.

10

11. A method according to claim 10, wherein the micelle-restoring material is  $\text{CaCl}_2$ .

12. A method according to any preceding claim, wherein the fluid is  
15 preconditioned to a pH in the range 6.5-7.5 before adding the alcohol.

13. A method according to any preceding claim, wherein the alcohol added to the fluid in an amount sufficient to produce an alcohol concentration, based on the total volume of fluid, of from 40-80% (v/v).

20

14. A method according to any preceding claim, wherein the alcohol added to the fluid in an amount sufficient to produce an alcohol concentration, based on the total volume of fluid, of from 50-70% (v/v).

25 15. A method according to any preceding claim, wherein the alcohol added to the fluid in an amount sufficient to produce an alcohol concentration, based on the total volume of fluid, of from 55-65% (v/v).

16. A method according to any preceding claim, wherein the alcohol added to  
30 the colostrum is substantially 100% pure.

17. A method according to any preceding claim, wherein the peptides recovered from the colostrum comprise colostrinin.

5

18. A method according to any preceding claim, wherein the fluid is a derivative of mammalian colostrum which has been subjected to one or more treatment steps.

10

Fig. 1.

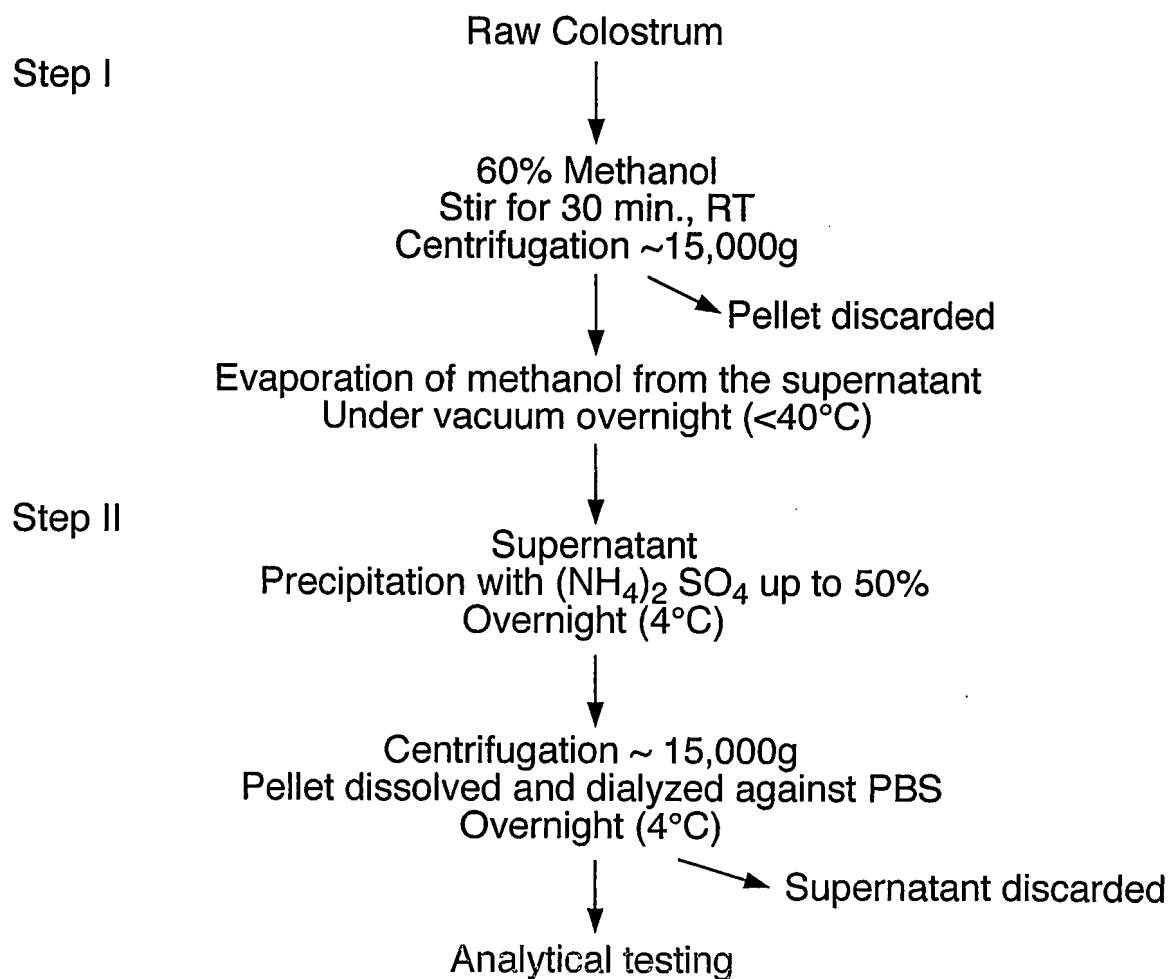
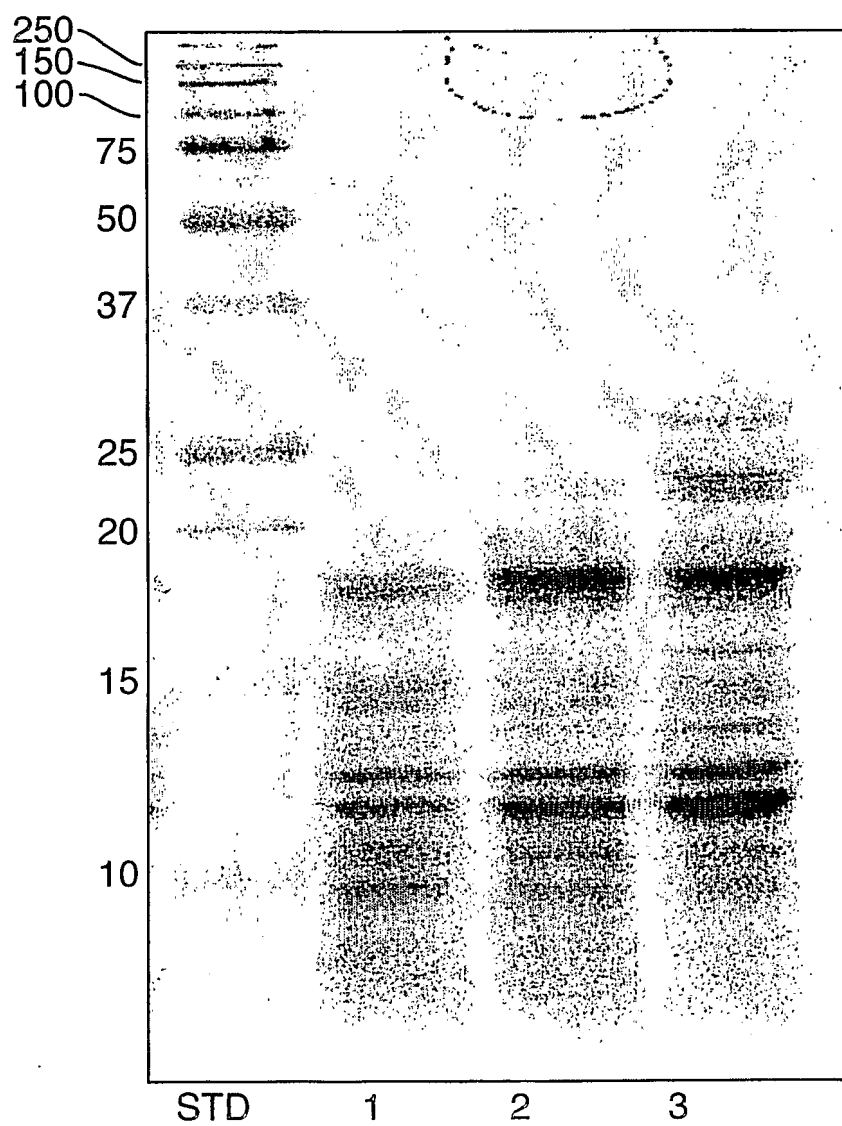




Fig.2.



20362WO.ST25.txt  
SEQUENCE LISTING

<110> ReGen Therapeutics plc  
 <120> Purification of Peptides from Colostrinin  
 <130> PAC/20362WO  
 <150> GB0305552.2  
 <151> 2003-03-11  
 <160> 64  
 <170> PatentIn version 3.1  
 <210> 1  
 <211> 17  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin  
 <400> 1

Leu Gln Thr Pro Gln Pro Leu Leu Gln Val Met Met Glu Pro Gln Gly  
 1 5 10 15

Asp

<210> 2  
 <211> 11  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin  
 <400> 2

Met Pro Gln Asn Phe Tyr Lys Leu Pro Gln Met  
 1 5 10

<210> 3  
 <211> 15  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin  
 <400> 3

Val Leu Glu Met Lys Phe Pro Pro Pro Pro Gln Glu Thr Val Thr  
 1 5 10 15

<210> 4  
 <211> 15  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin  
 <400> 4

Leu Lys Pro Phe Pro Lys Leu Lys Val Glu Val Phe Pro Phe Pro  
 1 5 10 15

<210> 5  
 <211> 4  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

20362WO.ST25.txt

&lt;400&gt; 5

Ser Glu Gln Pro

1

&lt;210&gt; 6

&lt;211&gt; 3

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 6

Asp Lys Glu

1

&lt;210&gt; 7

&lt;211&gt; 7

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 7

Asp Pro Pro Pro Pro Gln Ser

1

5

&lt;210&gt; 8

&lt;211&gt; 3

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 8

Leu Asn Phe

1

&lt;210&gt; 9

&lt;211&gt; 7

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 9

Val Leu Pro Pro Asn Val Gly

1

5

&lt;210&gt; 10

&lt;211&gt; 7

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 10

Lys Tyr Lys Leu Gln Pro Glu

1

5

&lt;210&gt; 11

&lt;211&gt; 5

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 11

20362WO.ST25.txt

Ser Glu Glu Met Pro  
1 5

<210> 12  
<211> 6  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 12

Asp Ser Gln Pro Pro Val  
1 5

<210> 13  
<211> 5  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 13

Phe Pro Pro Pro Lys  
1 5

<210> 14  
<211> 5  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 14

Val Val Met Glu Val  
1 5

<210> 15  
<211> 15  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 15

Asp Leu Glu Met Pro Val Leu Pro Val Glu Pro Phe Pro Phe Val  
1 5 10 15

<210> 16  
<211> 12  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 16

Leu Phe Phe Phe Leu Pro Val Val Asn Val Leu Pro  
1 5 10

<210> 17  
<211> 7  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 17

Met Gln Pro Pro Pro Leu Pro  
1 5

20362WO.ST25.txt

<210> 18  
 <211> 18  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

<400> 18

Asp Gln Pro Pro Asp Val Glu Lys Pro Asp Leu Gln Pro Phe Gln Val  
 1 5 10 15

Gln Ser

<210> 19  
 <211> 10  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

<400> 19

Val Tyr Pro Phe Thr Gly Pro Ile Pro Asn  
 1 5 10

<210> 20  
 <211> 9  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

<400> 20

Ser Leu Pro Gln Asn Ile Leu Pro Leu  
 1 5

<210> 21  
 <211> 10  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

<400> 21

Thr Gln Thr Pro Val Val Val Pro Pro Phe  
 1 5 10

<210> 22  
 <211> 18  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

<400> 22

Leu Gln Pro Glu Ile Met Gly Val Pro Lys Val Lys Glu Thr Met Val  
 1 5 10 15

Pro Lys

<210> 23  
 <211> 18  
 <212> PRT

20362WO.ST25.txt

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 23

His	Lys	Glu	Met	Pro	Phe	Pro	Lys	Tyr	Pro	Val	Glu	Pro	Phe	Thr	Glu
1				5					10					15	

Ser Gln

&lt;210&gt; 24

&lt;211&gt; 18

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 24

Ser	Leu	Thr	Leu	Thr	Asp	Val	Glu	Lys	Leu	His	Leu	Pro	Leu	Pro	Leu
1				5					10					15	

Val Gln

&lt;210&gt; 25

&lt;211&gt; 7

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 25

Ser	Trp	Met	His	Gln	Pro	Pro
1				5		

&lt;210&gt; 26

&lt;211&gt; 10

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 26

Gln	Pro	Leu	Pro	Pro	Thr	Val	Met	Phe	Pro
1				5					10

&lt;210&gt; 27

&lt;211&gt; 15

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 27

Met	His	Gln	Pro	Pro	Gln	Pro	Leu	Pro	Pro	Thr	Val	Met	Phe	Pro
1				5					10					15

&lt;210&gt; 28

&lt;211&gt; 6

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 28

Pro Gln Ser Val Leu Ser

20362WO.ST25.txt

1

5

<210> 29  
 <211> 22  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

<400> 29

Leu Ser Gln Pro Lys Val Leu Pro Val Pro Gln Lys Ala Val Pro Gln  
 1 5 10 15

Arg Asp Met Pro Ile Gln  
 20

<210> 30  
 <211> 7  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

<400> 30

Ala Phe Leu Leu Tyr Gln Glu  
 1 5

<210> 31  
 <211> 13  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

<400> 31

Phe Leu Leu Tyr Gln Glu Pro Val Leu Gly Pro Val Arg  
 1 5 10

<210> 32  
 <211> 8  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

<400> 32

Arg Gly Pro Phe Pro Ile Leu Val  
 1 5

<210> 33  
 <211> 18  
 <212> PRT  
 <213> Peptides from Ovine Colostrinin

<400> 33

Ala Thr Phe Asn Arg Tyr Gln Asp Asp His Gly Glu Glu Ile Leu Lys  
 1 5 10 15

Ser Leu

<210> 34  
 <211> 19

20362WO.ST25.txt

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 34

Leu Val Tyr Pro Phe Thr Gly Pro Ile Pro Asn Ser Leu Pro Gln Asn  
 1 5 10 15

Ile Leu Pro

&lt;210&gt; 35

&lt;211&gt; 13

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 35

Met Ile Val Val Arg Leu Leu Gln Asn Glu Val Pro Glu  
 1 5 10

&lt;210&gt; 36

&lt;211&gt; 10

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 36

Ser Leu Ser Gln Ser Lys Val Leu Pro Val  
 1 5 10

&lt;210&gt; 37

&lt;211&gt; 8

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 37

Leu Gln Thr Gln Thr Pro Val Val  
 1 5

&lt;210&gt; 38

&lt;211&gt; 7

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 38

Glu Met Pro Phe Pro Lys Tyr  
 1 5

&lt;210&gt; 39

&lt;211&gt; 6

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 39

Pro Val Glu Pro Phe Thr  
 1 5



20362WO.ST25.txt

<210> 40  
<211> 6  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 40

Val Pro Pro Phe Leu Gln  
1 5

<210> 41  
<211> 5  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 41

Pro Met Phe Leu Gln  
1 5

<210> 42  
<211> 5  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 42

Glu His Met Phe Val  
1 5

<210> 43  
<211> 4  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 43

Thr Asp Arg Asp  
1

<210> 44  
<211> 4  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 44

Val Gln Pro Thr  
1

<210> 45  
<211> 4  
<212> PRT  
<213> Peptides from Ovine Colostrinin

<400> 45

Pro Lys Val Lys  
1

<210> 46  
<211> 4

20362WO.ST25.txt

<212> PRT  
<213> Peptides from Ovine Colostrinin  
  
<400> 46

Asp Asp Asp Glu  
1

<210> 47  
<211> 4  
<212> PRT  
<213> Peptides from Ovine Colostrinin  
  
<400> 47

Thr Glu Glu Val  
1

<210> 48  
<211> 4  
<212> PRT  
<213> Peptides from Ovine Colostrinin  
  
<400> 48

Tyr Gln Gln Glu  
1

<210> 49  
<211> 4  
<212> PRT  
<213> Peptides from Ovine Colostrinin  
  
<400> 49

Phe Pro Pro Gln  
1

<210> 50  
<211> 4  
<212> PRT  
<213> Peptides from Ovine Colostrinin  
  
<400> 50

Gly Phe Gly Ile  
1

<210> 51  
<211> 3  
<212> PRT  
<213> Peptides from Ovine Colostrinin  
  
<400> 51

Leu Gln Ser  
1

<210> 52  
<211> 3  
<212> PRT  
<213> Peptides from Ovine Colostrinin

20362WO.ST25.txt

&lt;400&gt; 52

Val Val Val  
1

&lt;210&gt; 53

&lt;211&gt; 3

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 53

Gly Gly Lys  
1

&lt;210&gt; 54

&lt;211&gt; 3

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 54

Asp Met Val  
1

&lt;210&gt; 55

&lt;211&gt; 3

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 55

Glu Ser Gln  
1

&lt;210&gt; 56

&lt;211&gt; 3

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 56

Gly Arg Val  
1

&lt;210&gt; 57

&lt;211&gt; 3

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 57

Val Glu Glu  
1

&lt;210&gt; 58

&lt;211&gt; 3

&lt;212&gt; PRT

&lt;213&gt; Peptides from Ovine Colostrinin

&lt;400&gt; 58

20362WO.ST25.txt

Ile Gly Asn

1

<210> 59  
<211> 3  
<212> PRT  
<213> Peptides from Ovine Colostrinin

&lt;400&gt; 59

Phe Phe Gln

1

<210> 60  
<211> 3  
<212> PRT  
<213> Peptides from Ovine Colostrinin

&lt;400&gt; 60

Arg Met Phe

1

<210> 61  
<211> 3  
<212> PRT  
<213> Peptides from Ovine Colostrinin

&lt;400&gt; 61

Phe Pro Pro

1

<210> 62  
<211> 3  
<212> PRT  
<213> Peptides from Ovine Colostrinin

&lt;400&gt; 62

Met His His

1

<210> 63  
<211> 3  
<212> PRT  
<213> Peptides from Ovine Colostrinin

&lt;400&gt; 63

Asn Thr Glu

1

<210> 64  
<211> 9  
<212> PRT  
<213> Peptides from Ovine Colostrinin

&lt;400&gt; 64

Val Glu Ser Tyr Val Pro Leu Phe Pro

1

5

20362WO.ST25.txt

# INTERNATIONAL SEARCH REPORT

Int ional Application No  
PC1/GB2004/001014

**A. CLASSIFICATION OF SUBJECT MATTER**  
IPC 7 C07K14/47

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)  
IPC 7 C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, PAJ, WPI Data, BIOSIS

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2 890 986 A (RUTH KORBEL ET AL) 16 June 1959 (1959-06-16) examples 1,6	1-18
X	US 4 816 563 A (PADDOCK GARY V ET AL) 28 March 1989 (1989-03-28) cited in the application example 4	1-18
A	JANUSZ M ET AL: "CHEMICAL AND PHYSICAL CHARACTERIZATION OF A PROLINE-RICH POLYPEPTIDE FROM SHEEP COLOSTRUM" BIOCHEMICAL JOURNAL, PORTLAND PRESS, LONDON, GB, vol. 199, 1981, pages 9-15, XP002938312 ISSN: 0264-6021 page 10, left-hand column	

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

\* Special categories of cited documents :

\*A\* document defining the general state of the art which is not considered to be of particular relevance

\*E\* earlier document but published on or after the international filing date

\*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

\*O\* document referring to an oral disclosure, use, exhibition or other means

\*P\* document published prior to the international filing date but later than the priority date claimed

\*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

\*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

\*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

\*Z\* document member of the same patent family

Date of the actual completion of the international search

18 June 2004

Date of mailing of the international search report

23/07/2004

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,  
Fax: (+31-70) 340-3016

Authorized officer

Schmidt, Harald

# INTERNATIONAL SEARCH REPORT

Int ional Application No  
PCT/GB2004/001014

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	FR 2 617 049 A (HUET FRANCOIS) 30 December 1988 (1988-12-30) page 7, line 32 - page 9, line 18 -----	
A	PATENT ABSTRACTS OF JAPAN vol. 2000, no. 26, 1 July 2002 (2002-07-01) & JP 2001 240599 A (SNOW BRAND MILK PROD CO LTD), 4 September 2001 (2001-09-04) abstract -----	
A	WO 98/14473 A (JANUSZ MARIN ; LISOWSKI JOZEF (PL); DUBOWSKA INGLOT ANNA (PL); HIRSZFE) 9 April 1998 (1998-04-09) cited in the application claim 48; example 1 -----	
P,X	KRUZEL ML ET AL.: "The Alcohol-Induced Conformational Changes in Casein Micelles: A New Challenge for the Purification of Colostrinin" THE PROTEIN JOURNAL, vol. 23, no. 2, 2 February 2004 (2004-02-02), pages 127-133, XP008031778 page 129 - page 130; figure 3 -----	1-18

# INTERNATIONAL SEARCH REPORT

PCT/GB2004/001014

## Box No. 1 Nucleotide and/or amino acid sequence(s) (Continuation of item 1.b of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application and necessary to the claimed invention, the international search was carried out on the basis of:
  - a. type of material
    - ☒ a sequence listing
    - ☐ table(s) related to the sequence listing
  - b. format of material
    - ☒ in written format
    - ☒ in computer readable form
  - c. time of filing/furnishing
    - ☒ contained in the international application as filed
    - ☒ filed together with the international application in computer readable form
    - ☐ furnished subsequently to this Authority for the purpose of search
2. ☐ In addition, in the case that more than one version or copy of a sequence listing and/or table relating thereto has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that in the application as filed or does not go beyond the application as filed, as appropriate, were furnished.
3. Additional comments:



# INTERNATIONAL SEARCH REPORT

Information on patent family members

In International Application No

FL 1/GB2004/001014

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
US 2890986	A	16-06-1959	CH 338927 A	15-06-1959
			DE 950959 C	18-10-1956
			NL 89987 C	
			NL 197650 A	
<hr/>				
US 4816563	A	28-03-1989	AU 3577884 A	30-05-1985
			DK 558684 A	26-05-1985
			EP 0143445 A2	05-06-1985
			ES 8604021 A1	01-06-1986
			FI 844606 A	26-05-1985
			PT 79549 A ,B	01-12-1984
			JP 60208919 A	21-10-1985
			ZA 8409156 A	31-07-1985
<hr/>				
FR 2617049	A	30-12-1988	FR 2617049 A1	30-12-1988
<hr/>				
JP 2001240599	A	04-09-2001	NONE	
<hr/>				
WO 9814473	A	09-04-1998	PL 316416 A1	14-04-1998
			AU 4565197 A	24-04-1998
			BR 9712259 A	25-01-2000
			CA 2266859 A1	09-04-1998
			CN 1238782 A ,B	15-12-1999
			EP 0932623 A1	04-08-1999
			GB 2352176 A ,B	24-01-2001
			WO 9814473 A1	09-04-1998
			GB 2333453 A ,B	28-07-1999
			HU 9904368 A2	28-06-2000
			JP 2001501929 T	13-02-2001
			KR 2000048886 A	25-07-2000
			NZ 510681 A	20-12-2002
			PL 332632 A1	27-09-1999
			RU 2217152 C2	27-11-2003
			TR 9901022 T2	21-07-1999
			ZA 9708885 A	26-07-1999